

# The University of Chicago Genetic Services Laboratories



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## **EBP analysis for X-linked chondrodysplasia punctata**

### **Clinical Features:**

Patients with X-linked chondrodysplasia punctata (CDPX2) [OMIM #302960], also known as Happle syndrome or Conradi-Hünermann syndrome, have asymmetric shortening of the limbs, scoliosis, and widespread epiphyseal stippling, usually including the vertebral column and tracheal cartilage. Another classic finding includes various skin abnormalities, like erythema and scaling “oat bran” ichthyosis in the newborn period or atrophoderma and ichthyosis in older children. Congenital cataracts, microphthalmia, polydactyly, cleft palate, and visceral abnormalities have also been reported. Intelligence is usually normal. Severity in females varies greatly, from stillborns to females with very mild or unnoticeable symptoms (1).

Suggested minimal clinical criteria include **one or more of the following, along with increased levels of 8(9)-cholestenol:**

- scaling ichthyosis
- atrophoderma
- chondrodysplasia punctata on x-rays in infancy
- cataracts
- alopecia

### **Molecular and Biochemical Genetics:**

Mutations of the *EBP* [OMIM #300205] gene, or emopamil binding protein, have been identified in patients with CDPX2 (2, 3). *EBP* has 4 coding exons, and more than 55 mutations have been identified. No clear genotype-phenotype correlations have been reported, most likely due to random X-inactivation. This sterol- $\Delta^8$ - $\Delta^7$ -isomerase gene is the human homolog of *tattered* (*Td*) in mice. Affected hemizygous *Td* male mice die prenatally, and affected heterozygous *Td* female mice are dwarfed, exhibit hyperkeratotic eruption very early in life that resolves, and have similar biochemical findings as heterozygous *EBP* humans (1).

Patients with CDPX2 have increased tissue or plasma levels of 8(9)-cholestenol and 8-dehydrocholesterol. Sterol analysis of plasma and scales from skin lesions is currently used for diagnosis and is available at the Clinical Mass Spectrometry Laboratory at Kennedy Krieger Institute. This test may also distinguish CDPX2 from CHILD syndrome, a phenotypically similar condition caused by mutations in the *NSDHL* (NADH steroid dehydrogenase-like) gene (1). Unpublished data in Dr. Richard Kelley’s lab shows that approximately 95% of patients with these biochemical findings are found to have a mutation in the *EBP* gene.

### **Inheritance:**

CDPX2 is an X-linked condition that occurs in approximately 1 in 100,000 live births. CDPX2 is hypothesized to be lethal in most males, although a few affected hemizygous males with hypomorphic *EBP* mutations have been reported. Penetrance appears to be 100%, and incidence does not vary between populations. Germline mosaicism and/or somatic mosaicism have been reported (1). Recurrence risk for affected individuals and carrier mothers is 50%.

### **Test methods:**

We offer mutation analysis of all 4 coding exons and intron/exon boundaries of *EBP* by direct sequencing of amplification products in both the forward and reverse directions. We also offer oligonucleotide array-CGH analysis to identify deletions/duplications involving the coding region of the *EBP* gene. Deletions and duplications of less than 2 kb may not be detected by this methodology. Array-CGH will not detect low level mosaicism, balanced translocations, inversions, or point mutations that may be responsible for the clinical phenotype. The sensitivity of this assay may be reduced when DNA is extracted by an outside laboratory.

Patients with negative or unknown results can enroll in Dr. Aida Metzberg's research study (aida.metzenberg@csun.edu) at the California State University, Northridge for further studies.

*Please, send a completed CDPX2 Clinical Questionnaire and patient consent form with each sample.*

This information will be used to aid in interpretation of the test result. With the family's consent, the clinical data form along with the test result will be shared with Dr. Metzberg and entered anonymously into a database for research purposes.

**EBP sequencing analysis**

Sample specifications:	3 to10 cc of blood in a purple top (EDTA) tube
Cost:	\$540
CPT codes:	81404
Turn-around time:	4 - 6 weeks

**EBP deletion/duplication analysis**

Sample specifications:	3 to10 cc of blood in a purple top (EDTA) tube
Cost:	\$1000
CPT codes:	81403
Turn-around time:	4 weeks

**Testing for a known mutation in additional family members by sequence analysis**

Sample specifications:	3 to10 cc of blood in a purple top (EDTA) tube
Cost:	\$390
CPT codes:	81403
Turn-around time:	3-4 weeks

**Prenatal testing for a known mutation by sequence analysis**

Sample specifications:	2 T25 flasks of cultured cells from amniocentesis or CVS or 10 mL of amniotic fluid
Cost:	\$540
CPT codes:	81403
Turn-around time:	1-2 weeks

**Results:**

Results, along with an interpretive report, will be faxed to the referring physician. Additional reports will be provided as requested. All abnormal results will be reported by telephone.

**References:**

1. Herman GE. Disorders of cholesterol biosynthesis: prototypic metabolic malformation syndromes. Hum Mol Genet 2003; 12 Spec No 1: R75-88.
2. Derry JM, Gormally E, Means GD et al. Mutations in a delta 8-delta 7 sterol isomerase in the tattered mouse and X-linked dominant chondrodysplasia punctata. jderry@immunex.com. Nat Genet 1999; 22: 286-290.
3. Braverman N, Lin P, Moebius FF et al. Mutations in the gene encoding 3 beta-hydroxysteroid-delta 8, delta 7-isomerase cause X-linked dominant Conradi-Hünemann syndrome. Nat Genet 1999; 22: 291-294.

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